# MYCOTAXON

Volume 115, pp. 121-124

DOI: 10.5248/115.121

January–March 2011

## First report of Conidiobolus coronatus in Turkey

CAFER EKEN<sup>1,2\*</sup>, Şaban Güçlü<sup>2</sup> & Kibar Ak<sup>3</sup>

<sup>1</sup>Graduate School of Natural and Applied Sciences, Ardahan University, Ardahan,75000,Turkey <sup>2</sup>Department of Plant Protection, Faculty of Agriculture, Atatürk University, Erzurum,25240, Turkey <sup>3</sup>Black Sea Agricultural Research Institute, Gelemen, Samsun,55001, Turkey \*CORRESPONDENCE TO: cafereken@hotmail.com

ABSTRACT — *Conidiobolus coronatus (Entomophthorales, Zygomycota)* was isolated from infected specimens representing an *Issus* sp. (*Issidae, Hemiptera*) collected from the Trabzon province of Turkey. The species, which represents a new record for the Turkish mycoflora, is described briefly and illustrated.

KEY WORDS — insect, entomopathogenic fungi, hazelnut

#### Introduction

Members of the widespread order *Entomophthorales* (*Zygomycota*) are predominantly pathogens of insects and mites (Pell et al. 2001). Species of *Conidiobolus*, most notably the ubiquitous *C. coronatus*, are recorded as widespread soil saprophytes utilising a variety of substrates, including plant detritus, living plants, different dead arthropods and the fruiting bodies of other fungi in various regions of the world (MacLeod & Müller-Kögler 1973, Keller 1987, Sajap et al. 1997, Dromph et al. 2001, Laxman et al. 2005, Manning et al. 2007, Comerio et al. 2008). *Conidiobolus coronatus*, mainly tropical strains, is known to cause disease in both insects and humans (Ribes et al. 2000, Prabhu & Patel 2004); the disease has been named rhinophycomycosis, rhinophycomycosis entomophthorae, rhinoentomophthoromycosis, and conidiobolomycosis (King 1979, Ochoa et al. 1996, Yang et al. 2010).

#### Material & methods

Dead insects collected in 2008 from a hazelnut orchard in in the Black Sea region of Turkey were cultured for entomopathogenic fungi. After the cadavers were washed in a solution of 2% sodium hypochlorite for 1 min, they were dried on filter paper. After transfer to Petri dishes containing 20 ml of PDA, the cadavers were incubated at 25°C for 1 week with high humidity (80  $\pm$  10% rh). Colonies of filamentous fungi

#### 122 ... Eken, Güçlü & Ak

emerging from each cadaver and identifiable as the genus *Conidiobolus* were transferred to PDA and identified to species using the relevant literature (Emmons & Bridges 1961, Prasertphon 1963, MacLeod & Müller-Kögler 1973, King 1979, Keller 1987, Humber 1997, Hatting et al. 1999, Toledo et al. 2007, Comerio et al. 2008). After identification, all isolates were deposited in the fungal collection of Department of Plant Protection, Faculty of Agriculture, Atatürk University, Erzurum-Turkey.

## Results

The description and illustration of *Conidiobolus coronatus* given below are based on the Turkish collections of the material. This is the first report of *C. coronatus* from Turkey.

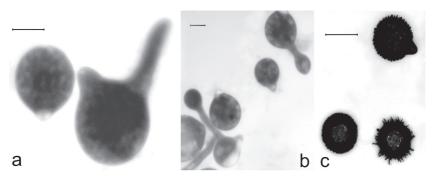


FIGURE 1. *Conidiobolus coronatus*: A— primary conidium and germinating primary conidium; B— primary conidia and secondary conidia produced on germ tube arising from primary conidia; C— villose conidia may be the equivalent of resting spores in this species. Bar = 20 μm.

### Conidiobolus coronatus (Costantin) A. Batko, Entomophaga, Mem. Hors, Ser. 2: 129, 1964.

FIG. 1

COLONIES (PDA) expanding, hyaline, soon with irregular radial blooms; at 25°C reaching 85 mm diam. after 2 days; PRIMARY CONIDIA (FIG. 1A) globose, with more prominent basal papilla tapering toward obtuse apex; variable in size,  $(23.2-)32.1(-46.8) \times (18.1-)24.6(-35.1) \mu$ m; basal papilla prominent with pointed apex, forcibly discharged; SECONDARY CONIDIA (FIG. 1B) forming singly and forcibly discharged or (more commonly) producing many forcibly discharged secondary microconidia on short germ tubes arising from primary conidia; CONIDIOPHORES simple, unbranched; VILLOSE CONIDIA (resting spores; FIG. 1C) resembling primary conidia but covered with villose appendages (unique to this species), (19.9–)25.8(–34.8)  $\mu$ m diam.

SPECIMEN EXAMINED: On cadavers of *Issus* sp. (*Issidae, Hemiptera*). **TURKEY: TRABZON PROVINCE, Of district**, *Corylus avellana* L. (*Corylaceae*) orchards, 40°51'30"N, 40°16'00"E, alt. 160 m, VI-VIII 2008, coll. K AK (KA 123).

#### Discussion

*Conidiobolus coronatus*, originally isolated from a culture of *Agaricus campestris* L. (possibly derived from a dead insect hidden between the lamellae), was described in 1897 as *Boudierella coronata* Costantin (MacLeod & Müller-Kögler 1973). Since then it has been isolated from numerous and diverse sources (MacLeod & Müller-Kögler 1973, King 1979, Keller 1987, Sajap et al. 1997, Dromph et al. 2001, Laxman et al. 2005, Manning et al. 2007, Comerio et al. 2008). Utilization of the species as a biological control is limited by its potential to cause human disease (King 1979, Ochoa et al. 1996, Ribes et al. 2000, Prabhu & Patel 2004).

Diameters of both the primary and villose conidia measured in this study fell within the ranges observed by Emmons & Bridges (1961; primary conidia:  $36-44 \ \mu\text{m}$ ), Prasertphon (1963; primary:  $25-61 \ \mu\text{m}$ ; villose:  $8-42 \ \mu\text{m}$ ), Keller (1987; primary:  $37-74 \ \mu\text{m}$ ; villose:  $16-42 \ \mu\text{m}$ ), Hatting et al. (1999; primary:  $41.5-66 \times 30-48 \ \mu\text{m}$ ), Toledo et al. (2007; primary:  $17.6-39.5 \times 24.7-29.6 \ \mu\text{m}$ ; villose:  $19.8-24.7 \ \mu\text{m}$ ), and Comerio et al. (2008; primary:  $30-38 \ \mu\text{m}$ ).

#### Acknowledgments

We are grateful to Dr. Celal Tuncer and Dr. Yusuf Yanar for reviewing the manuscript.

#### Literature cited

- Comerio RM, Andorno AV, Botto EN. 2008. *Conidiobolus coronatus* isolation from a pest aphid of chives (*Allium schoenoprasum L.*). Revista Iberoamericana de Micología 25: 193–195.
- Dromph KM, Eilenberg J. Esbjerg P. 2001. Natural occurrence of entomophthoralean fungi pathogenic to collembolans. Journal of Invertebrate Pathology 78: 226–231. doi: 10.1006/ jipa.2002.5077
- Emmons CW, Bridges CH. 1961. Entomophthora coronata, the etiologic agent of a phycomycosis of horses. Mycologia 53: 307–312. doi: 10.2307/3756277
- Hatting JL, Humber RA, Poprawski TJ, Miller RM. 1999. A survey of fungal pathogens of aphids from South Africa, with special reference to cereal aphids. Biological Control 16: 1–12. doi: 10.1006/bcon.1999.0731
- Humber RA. 1997. Fungi: identification. 153–185, in LA Lacey (ed.), Manual of Techniques in Insect Pathology. London, Academic Press.
- Keller S. 1987. Arthropod-pathogenic Entomophthorales of Switzerland. I. Conidiobolus, Entomophaga and Entomophthora. Sydowia 40: 122–167.
- King DS. 1979. Systematics of fungi causing entomophthoramycosis. Mycologia 71: 731–745. doi: 10.2307/3759185
- Laxman RS, Sonawane AP, More SV, Rao BS, Rele MV, Jogdand VV, Deshpande VV, Rao MB. 2005. Optimization and scale up of production of alkaline protease from *Conidiobolus coronatus*. Process Biochemistry 40: 3152–3158. doi: 10.1016/j.procbio.2005.04.005
- MacLeod DM, Müller-Kögler E. 1973. Entomogenous fungi: Entomophthora species with pearshaped to almost spherical conidia (Entomophthorales: Entomophthoraceae). Mycologia 65: 823–893. doi: 10.2307/3758521

- Manning RJ, Waters SD, Callaghan AA. 2007. Saprotrophy of *Conidiobolus* and *Basidiobolus* in leaf litter. Mycological Research 111: 1437–1449. doi: 10.1016/j.mycres.2007.08.019
- Ochoa LF, Dubque CS, Velez A. 1996. Rhinoentomophthoromycosis. Report of two cases. Journal of Laryngology and Otology 110: 1154–1156. doi: 10.1017/S002221510013600X
- Pell JK, Eilenberg J, Hajek AE, Steinkraus DC. 2001. Biology, ecology and pest management potential of *Entomophthorales*. 71–153, in TM Butt et al. (eds.), Fungi as Biocontrol Agents. Wallingford, CABI Publishing. doi: 10.1079/9780851993560.0071
- Prabhu RM, Patel R. 2004. Mucormycosis and entomophthoramycosis: a review of the clinical manifestations, diagnosis and treatment. Clinical Microbiology and Infection 10 (Suppl 1): 31–47. doi: 10.1111/j.1470-9465.2004.00843.x
- Prasertphon S. 1963. Conidial formation in *Entomophthora coronata* (Costantin) Kevorkian. Journal Insect Pathology 5: 318–335.
- Ribes JA, Vanover-Sams CL, Baker DJ. 2000. Zygomycetes in human disease. Clinical Microbiology Reviews 13: 236–301. doi: 10.1128/CMR.13.2.236-301.2000
- Sajap AS, Atim AB, Husim H, Wahab YA. 1997. Isolation of Conidiobolus coronatus (Zygomycetes: Entomophthorales) from soil and its effect on Coptotermes curvignathus (Isoptera: Rhinotermitidae). Sociobiology 30: 257–262.
- Toledo AV, Remes Lenicov AMM de, López Lastra CC. 2007. Primer registro de Conidiobolus coronatus (Zygomycetes: Entomophthorales) en crías experimentales de dos especies plaga del maíz: Delphacodes kuscheli y D. haywardi (Hemiptera: Delphacidae) en la Argentina. Boletín de la Sociedad Argentina de Botánica 42: 169–174.
- Yang X, Li Y, Zhou X, Wang Y, Geng S, Liu H, Yang Q, Lu X, Hiruma M, Sugita T, Ikeda S, Ogawa H. 2010. Rhinofacial conidiobolomycosis caused by *Conidiobolus coronatus* in a Chinese rice farmer. Mycoses 53: 369–373. doi: 10.1111/j.1439-0507.2009.01716.x